

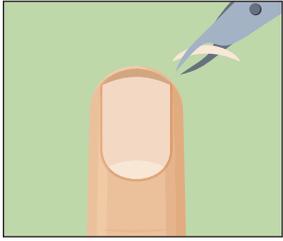
Assurance Scientific Laboratories offers a solution for your patients with suspected fungal infections (i.e. skin, nail, etc.). Utilizing PCR, our lab can identify the fungus causing the infection, which can help guide treatment. With a simple nail clipping or swab of the infected area, Assurance can test for 16 fungal pathogens. As part of our fungal infection panel, we also offer a bacterial add on panel to identify if *mecA* and *Pseudomonas aeruginosa* are present. Results are returned within 48 hours.

Custom fungal pathogens and other targets can be made for your needs. Talk to a representative for more details. Let us provide you with diagnostic assurance!

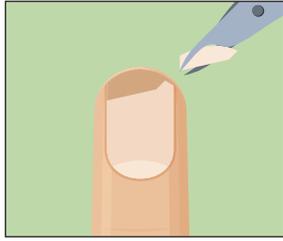
PATHOGENS
<i>Alternaria</i> spp.
<i>Aspergillus</i> spp.
<i>Fusarium</i> spp.
<i>Scytalidium dimidiatum</i>
<i>Sarocladium strictum</i>
<i>Candida albicans</i>
<i>Candida glabrata</i>
<i>Candida krusei</i>
<i>Candida parapsilosis</i>
<i>Candida tropicalis</i>
<i>Cryptococcus</i> spp.
<i>Malassezia</i> spp.
<i>Meyerozyma guilliermondii</i>
<i>Trichophyton anthropophilic</i> spp.
<i>Trichophyton zoophilic</i> spp.
<i>Microsporum canis</i>
<i>Trichosporon</i> spp.
<i>Epidermophyton floccosum</i>
<i>Curvularia</i> spp.
Bacterial Add On (includes ABX Resistant Marker)
<i>Pseudomonas aeruginosa</i>
Methicillin/Oxacillin ( <i>mecA</i> )

# FUNGAL INFECTION COLLECTION INSTRUCTIONS

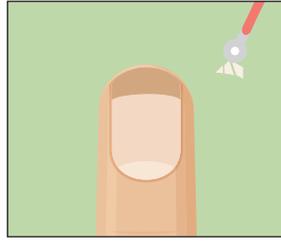
## Nail Clipping Instructions:



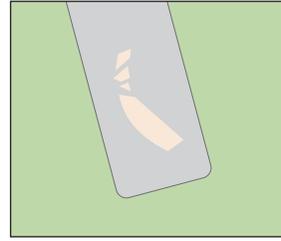
1. Wipe nail collection site with 70% isopropyl alcohol (local anesthesia may be required). Debride and discard distal nail clippings.



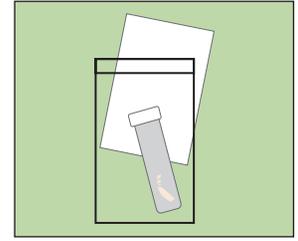
2. Obtain specimen from the most proximal area of nail and hyponychium. Minimum specimen amount of nail and subungual debris is 3mm to 6mm+ (*small pieces to obtain this size are preferred*).



3. Use curette to obtain additional subungual debris, as this will increase the potential yield.

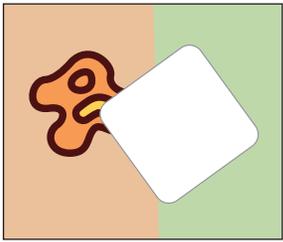


4. Place dry nail sample and subungual debris into a provided dry collection tube. Place the tube into a provided Assurance collection bag.

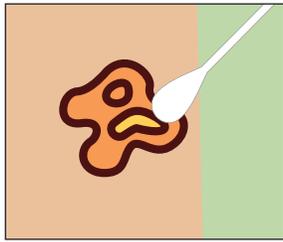


5. Complete information on the requisition form and attach the form to the collection bag.

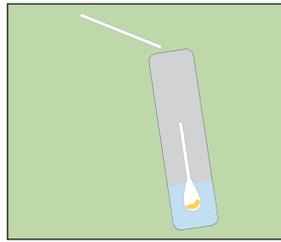
## Swab Instructions:



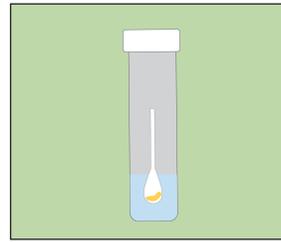
1. Cleanse and debride the infected area with 70% isopropyl alcohol using sterile gauze. Saline can be used if there is an open wound.



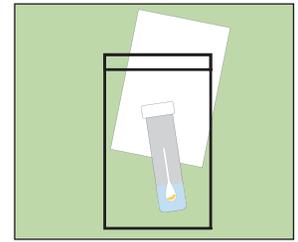
2. Exudate and brushings of the base of the infected area (*including advancing margins*) should be collected using the swab.



3. Without contaminating the swab, place the swab into the transport tube all the way to the bottom. Break the swab at the scored breakpoint. Leave the bottom portion inside the tube submerged in buffer solution.

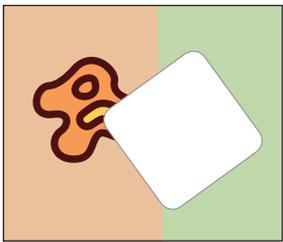


4. Secure the specimen by closing the tube lid tightly and placing it into a provided collection bag.

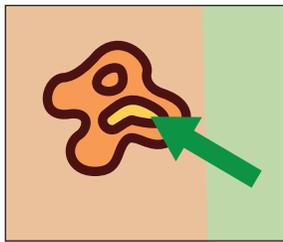


5. Complete information on the requisition form and attach the form to the collection bag.

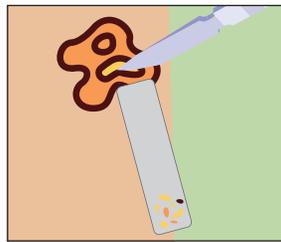
## Skin Scraping Instructions:



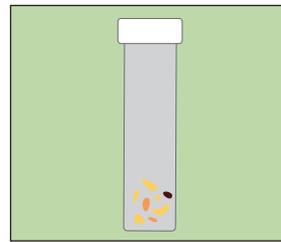
1. Remove any traces of skin products, medications, or surface contaminants by wiping the area with a 70% isopropyl alcohol wipe.



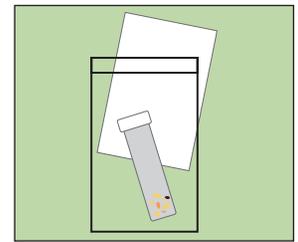
2. Choose the best area to scrape by determining the area where the fungal growth is most active.



3. Scrape the skin using a scalpel held at a blunt angle into a dry collection tube (*the greater the amount of specimen, the better the result*).



4. Secure the specimen by closing the tube lid tightly and placing it into a provided collection bag.



5. Complete information on the requisition form and attach the form to the collection bag.